

Active Participation of Dendritic Cells in Tumour Cell Immune Evasion

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Abstract

Conveying influences of a dual participation of dendritic cells and of tumour cells emerge in the shaping and reshaping of phenotype characterization of the high degree of plasticity attributes of dendritic cells (DC) in particular. In such terms, DC come to play an active essential role of the tumour cell evasion of the immune response that is transferable in terms of the contextual influences and modulations, as defined by immature versus mature stage specificities of the dendritic cells. In terms that go beyond such considerations, the incremental responses of intracellular mediators within both dendritic cells and tumour cells convey a realization that involves, in inherent manner, the evasion of the tumour cells from the immunosurveillance programs otherwise projected by dendritic cells.

1. Introduction

Signalling pathways in both tumour cells and dendritic cells (DC) constitute a predominant system series of inhibitors in suppressing immunosurveillance of tumours. In such manner, inhibitors of p38 MAPK, JAK/STAT3, PI3K/Akt and NF kappa constitute possible mechanisms as target molecular pathways in enhancing differentiation and functional activity in DC.

Transforming growth factor (TGF)-beta is central to immune suppression with roles in tutor immune evasion and poor responses to cancer immunotherapy [1].

The constitutive mechanistic pathways of tumour cells involve the secretion of various cytokines such as IL-6, IL-10, and TGFbeta, as after the exposure of DC to tumour culture-conditioned medium and inhibitors or antibodies against these specific cytokines that restore differentiation and functionality of DC against tumour cells. Natural Killer cells stimulate DC recruitment into the tumour microenvironment, enhancing cancer immune control; a cellular and molecular checkpoint for intratumoural DC recruitment is targeted by tumour-derived prostaglandin E2 for immune evasion [2].

2. Intracellular Pathways

Such considerations incorporate the essential roles of intracellular pathway mechanisms in the tumour cell evasion of immune response pathway dynamics of DC. Mechanisms are utilised by cancers to induce DC toleration, with various parallels between the evolution of these mechanisms and the process of mesenchymal transformation involved in tumourigenesis and metastasis [3].

In such terms, ongoing derivative dimensions of intracellular signalling may be transferable factors in the evolutionary history of tumour cell evasion from the immunosurveillance programs as conveyed by DC. The causes of breast cancer's immune silence derive from mechanisms that diminish immune recognition and others that promote strong immunosuppression [4].

An essential feature of tumour evasion is the effect of secreted soluble factors that tumour cells produce in the abrogation of the immune response as resulting from impaired differentiation and maturation of DC from bone marrow hematopoietic progenitor cells and monocytes. In such terms, ongoing system profile derivation is intrinsic historic role for the activation of antitumour immune responses.

The depiction of area and regional systems is conveyed especially by the localization of DC in peripheral tissues that reach regional lymph nodes in the differentiation programs undergone by circulating DC. It is significant to view DC participatory roles in immunosurveillance as target inhibition by evading tumour cells. New immune therapies can reverse immune evasion strategies of tumour cells, with also the blockade of immune checkpoints cytotoxic T-lymphocyte antigen 4 and programmed death-1 [5].

3. Stage Dependence

Stage dependence of p38 MAPK (Mitogen activated protein kinase) is illustrated by inhibition of DC differentiation and maturation when DC is derived from monocytes. The incremental dimensions of such stage-dependence is assumed within the context of the need for p38 MAPK in the generation of immature DC to form mature and cytokine-secreting DC. It is significant to recognize the plasticity of DC as conveyed by stage-dependence in the maturation of immunosurveillance programs, in response to a multitude of evading systems as transferable Microsystems carried forward by tumour cells. The programmed cell death protein 1 pathway plays a role in eliciting the immune checkpoint response of T cells and application of anti-PD-1/PD-L1 antibodies as checkpoint inhibitors are rapidly becoming a promising therapeutic approach [6].

It is further to such conditioned mediators produced by tumour cells that DC play also active roles in tumour evasion by means of manipulated maturational context as transferred by immature DC. In such terms, ongoing programs of interaction of immature DC and of actively proliferating cells include various intracellular signaling pathways that are stage dependent. Mechanisms have evolved in cancers to alter DC metabolic pathways, thus allowing for continued tumour progression and metastasis [7].

4. Myeloma Patients

In myeloma patients, mature DC produce significantly less CD1a, CD40, CD80 and HLA-DR and are poor in activating autologous antigen-specific T cells. Such phenomena are indicative of a transforming series of activities that are derived from interactivities of DC with tumour cells. The attributes of such interactivities between DC and tumour cells may be conveyed by elevated production of such autologous cytokines as represented by IL-6, activated p38 MAPK and STAT3, and inhibited MEK/ERK signalling pathways in DC progenitor cells.

The immunosuppressive tumour microenvironment is a major barrier to immunotherapy and tumour-derived retinoid acid regulates intratumour monocyte differentiation to enhance immune suppression [8].

5. Reprogramming

Attenuation of regulatory T cells in response to Toll-like receptor agonists results from inhibiting p38 MAPK signalling. Thus, inhibiting p38 MAPK by SB203580 allows for the emergence of promoted type 1 T cell responses. An understanding of the differential maturation steps of DC may help clarify the mechanisms of initial commitment of tumour cells that are involved in the emergence of either Th1 or Th2 subsets. ERK inhibition is also a potential target mechanism for contextual evasion of the tumour cells from the immune system response.

6. Dendritic Cell Maturation

JAK/STAT3 signaling pathways are also inhibiting systems in DC maturation, and as such, are instrumental in further conditioning the antitumour immune response. Cross-talk with the NF kappaB on the part of JAK/STAT, p38 MAPK and ERK signaling pathways potentiates the transcription programs of target genes, such as the expression of IL-12. A significant correlate is the high level of activity of NF kappaB in tumour-induced DC dysfunction in patients with cancer.

One essential mechanism behind CD47-mediated immune evasion is interaction with signals regulatory protein-alpha (SIRPalpha) expressed on myeloid cells; this induces phosphorylation of SIRPalpha cytoplasmic immunoreceptor tyrosinebased inhibition motifs and recruitment of Src homology 2 domain-containing tyrosine phosphatases to ultimately result in delivering an anti-phagocytic-signal [9].

7. Specifically High Levels of Signaling

High levels of signaling pathway mediators are a specific mechanism in the deregulating of maturation programs that result specifically in tumour evasion of the immune system responses. As such, hyperactivation of key signaling pathways is parent mechanism in the deregulation of the interacting dynamics of the DC with tumour cells, as these latter produce and maintain contextual evasion from the immune response.

Preclinical data show that inhibition of cyclooxygenase synergizes with anti-PD-1 blockade in inducing tumour eradication, thus suggesting that COX inhibitors may be useful adjuvants in immunotherapy of tumour patients [10,11].

Recent preclinical data suggest new therapies to subvert tutor induced immunosuppression via prostaglandin inhibition.

8. Concluding Remarks

A scenario of interactivities of DC with tumour cells emerges as terms of reference of intracellular signalling pathways in both DC and malignant cells.

REFERENCES

- 1. Batlle E, Massague J. Transforming Growth Factor-beta signaling in immunity and cancer. Immunity. 2019;50(4):924-40.
- 2. Bottcher JP, Bonavita E, Chakravarty P, et al. NK cells stimulate recruitment of cDC1 into the tumour microenvironment promoting cancer immune control. Cell. 2018;172(5):1022-37.
- 3. DeVito NC, Plebanek MP, Theivanthiran B, et al. Role of tumour-mediated dendritic cell tolerization in in immune evasion. Front Immune. 2019;10:2876.
- 4. Bates JP, Derakhshandeh R, Jones L, et al. Mechanisms of immune evasion in breast cancer. BMC Cancer. 2018;18(1):556.
- Muenst S, Laubli H, Soysal SD, et al. The immune system and cancer evasion strategies: therapeutic concepts. J Intern Med. 2016;279(6):541-62.
- Wu X, Gu Z, Chen Y, et al. Application of PD-1 blockade in cancer immunotherapy. Comput Struct Biotechnol J. 2019;17:661-74.
- 7. Plebanek MP, Sturdivant M, DeVito NC, et al. Role of dendritic cell metabolic reprogramming in tumour immune evasion. Int Immune. 2020;32(7):485-91.
- 8. Devalaraja S, To TKJ, Folkert IW, et al. Tumour-derived retinoic acid regulates intratumour monocyte differentiation to promote immune suppression. Cell. 2020;180(6):1098-1114.
- 9. Liu X, Kwon H, Li Z, et al. Is CD47 an innate immune checkpoint for tumour evasion? J Hematol Oncol. 2017;10(1):12.
- 10. Zelenay S, van der Veen AG, Bottcher JP, et al. Cycloosygenase-dependent tumour growth through evasion of immunity. Cell. 2015;162(6):1257-70.
- 11. Wang D, DuBois RN. The role of prostaglandin E(2) in tumour-associated immunosuppression. Trends Mol Med. 2016;22(1):1-3.